

Microdissection: Rationale and Applications

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Disclosure

- I DO NOT have any connection with the brand names, manufacturers and their representatives seen and/or mentioned in this presentation

Overview

- Definition
- Basic idea of microdissection
- History
- Types of microdissection
- Laser-assisted microdissection
- Available platforms
- Specimen preparation and expected yield of nucleic acids and proteins
- Selected applications in pathology and examples of downstream analyses
- Future perspectives



Definition

- Separation of cells or groups of cells having **specific features** from the “background” population

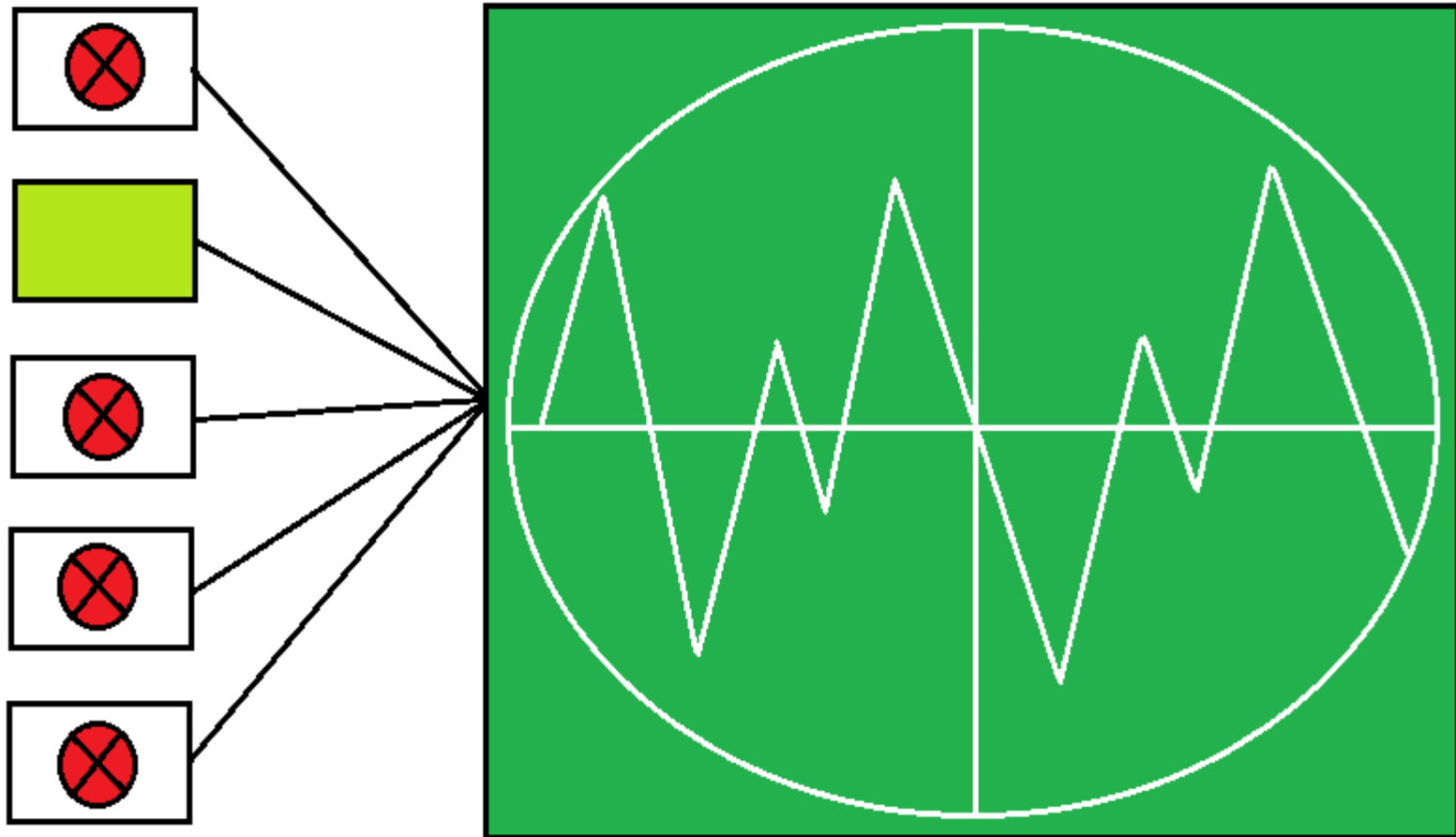
Definition

- These specific features can be:
 - Morphological properties only
 - Morphology + histochemistry / immunohistochemistry
 - Functional (biological behavior)

Definition

- Aim:
 - To obtain “pure” raw material for downstream analysis
 - To eliminate unwanted components from the section on the slide
 - To enrich selected types of cells for genomic or proteomic studies

Basic Idea of Microdissection



History

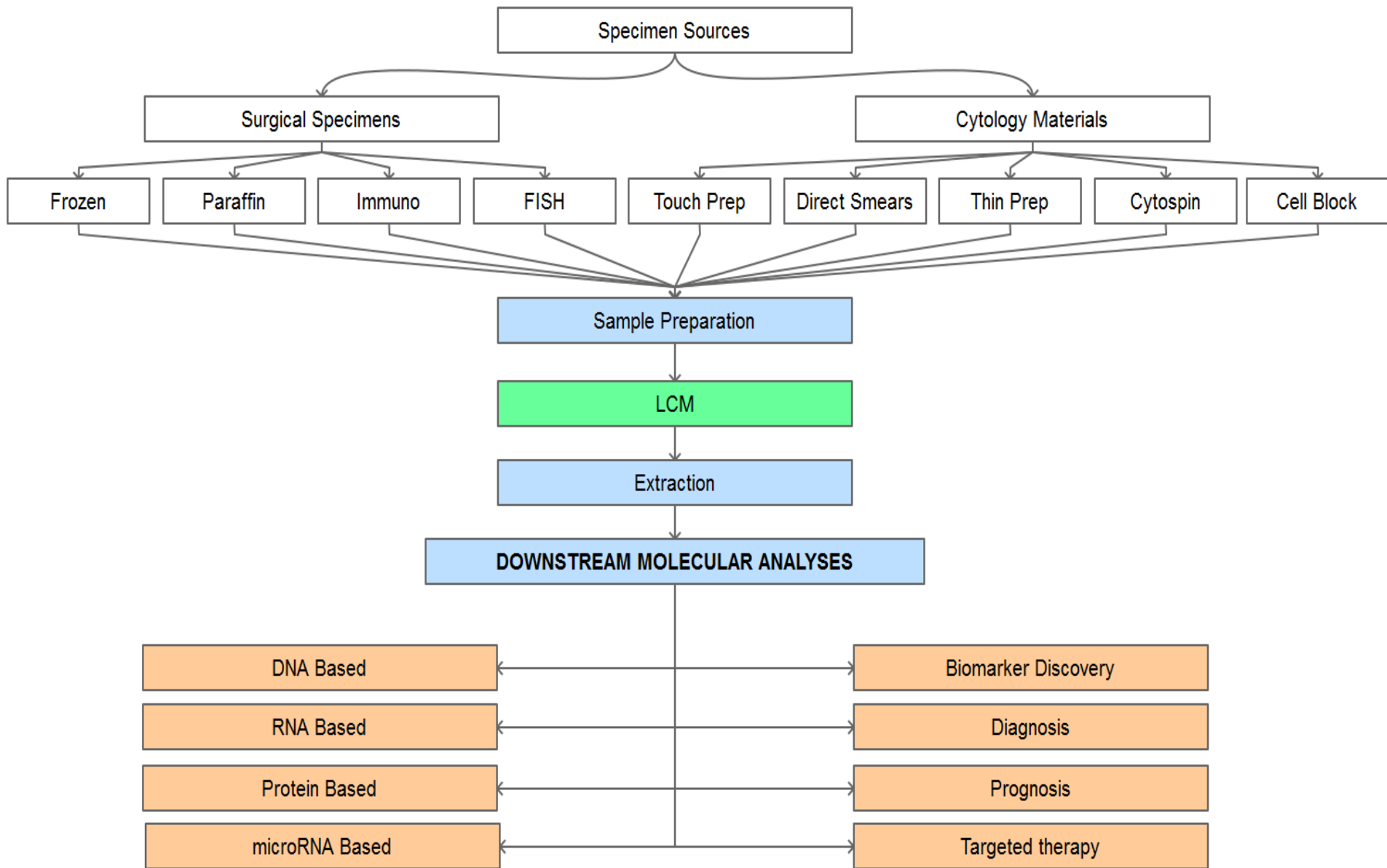
- The idea is not new
- First attempts were based on the usage of razor blades under a dissecting microscope
- Biochemical analysis on specific cell types from lyophilized tissues (Lowry and Passoneau, 1972)
- Manual extraction from formalin-fixed paraffin-embedded (FFPE) tissues by Goelz et al. (1985)

History

- The concept of using laser beam goes back 1969 (Berns-Rounds)
- First UV-laser equipped microscope in 1976 (Isenberg, Meier-Ruge)
- First prototypes of laser-based systems were massively space-occupying 😊
- First advanced laser-microdissection instrument in 1996 (Emmert-Buck)

History

- This prototype was commercialized by Arcturus Engineering
- Then, several competitors were appeared
- Most widely used systems are Laser Capture Microdissection instruments
- Several modifications and improvements became available on the growing market



Tissue Conditions

Sample Isolation

Analyte Amplification

Analysis

Markets

Healthy ↔ Diseased

Treated ↔ Untreated

Microdissection

DNA RNA Protein

Need > 500 / 2000 cells

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+

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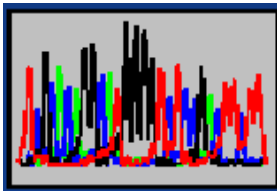
Structural Genomics

Functional Genomics (Biochips)

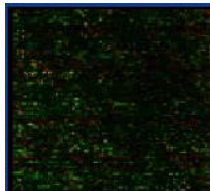
Genome

Transcriptome

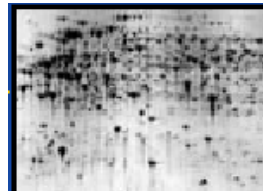
Proteome



- Genome mapping
- Genome sequencing
- Genome annotation



- DNA arrays and chips
- Real-time PCR
- Northern hybridization
- Transcriptional fusions



- 2D gel electrophoresis
- Mass spectrometry
- Protein sequencing

Molecular Pathology

Oncology

Neuroscience

Toxicogenomics

Pharmacogenomics

Neuroscience

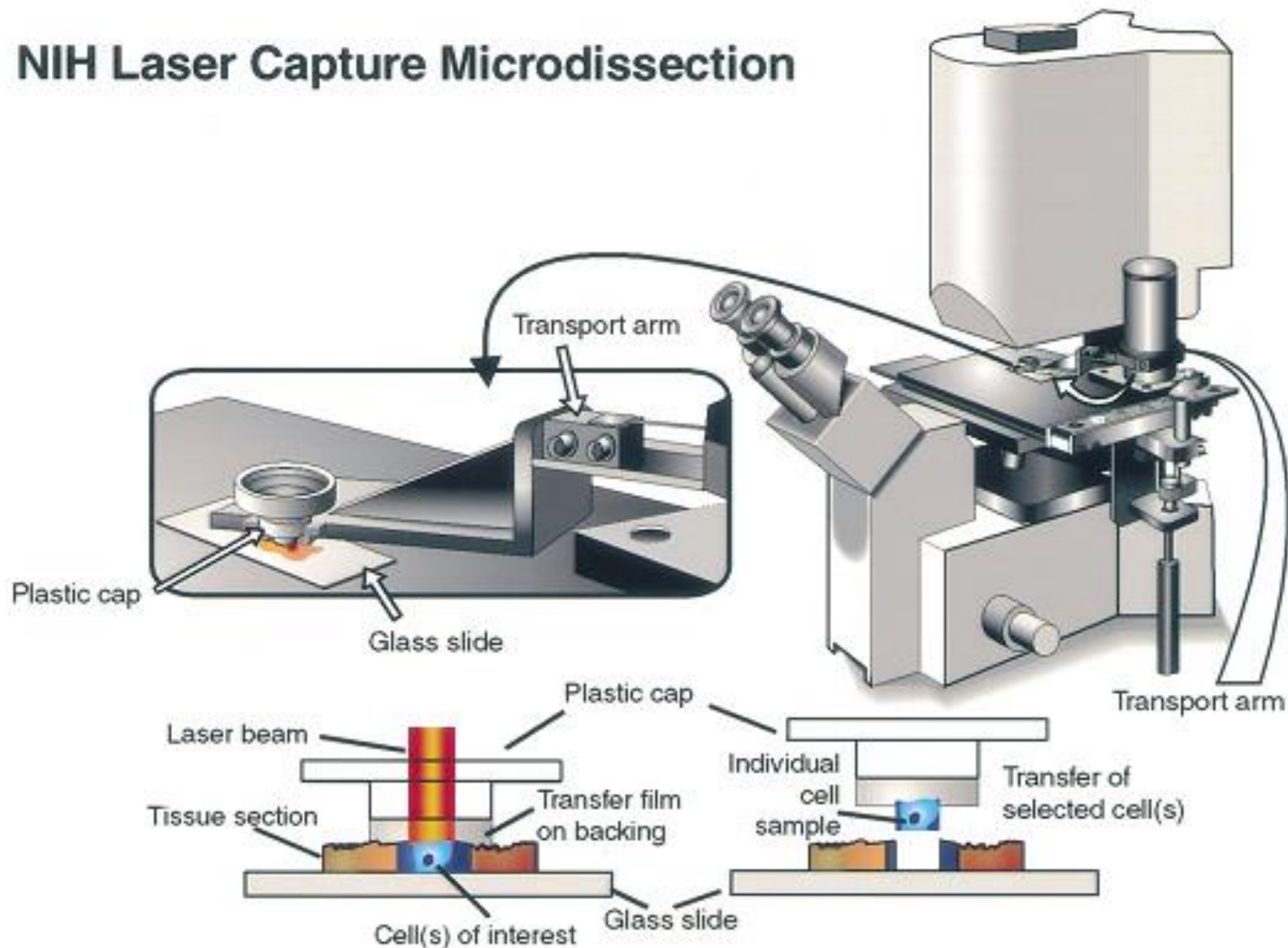
Types of Microdissection

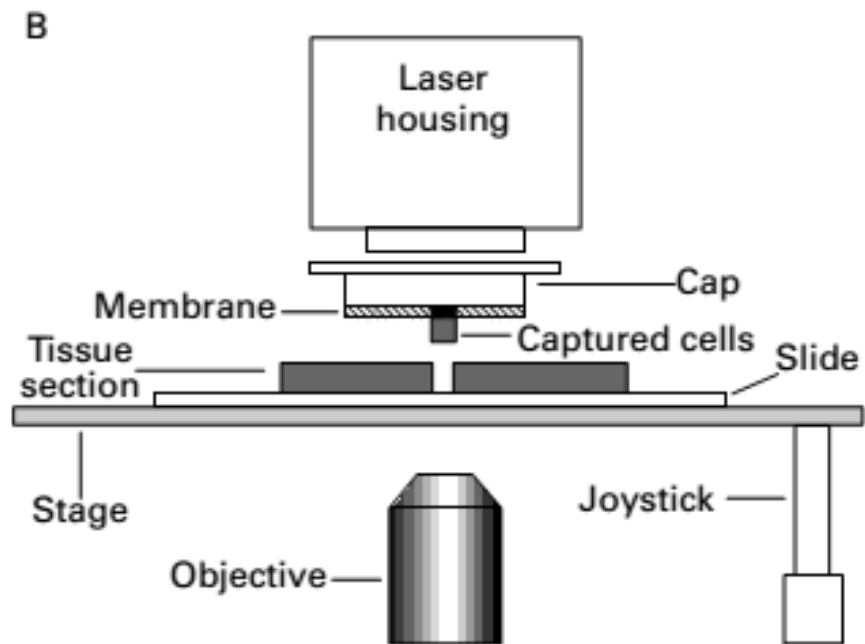
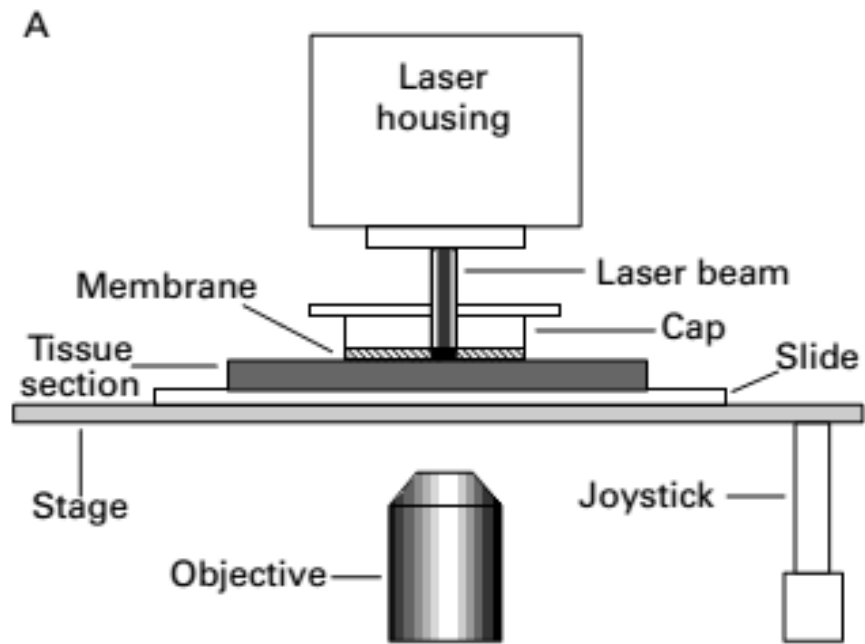
- Manual microdissection
 - Scalpels, razor blades, needle-tips
 - Dissection microscope + micromanipulator
- Instrument-assisted microdissection
 - Devices using needles with ultrasonic vibrations
 - **Laser capture microdissection**
 - **Laser cutting microdissection**

Laser-capture microdissection (LCM)

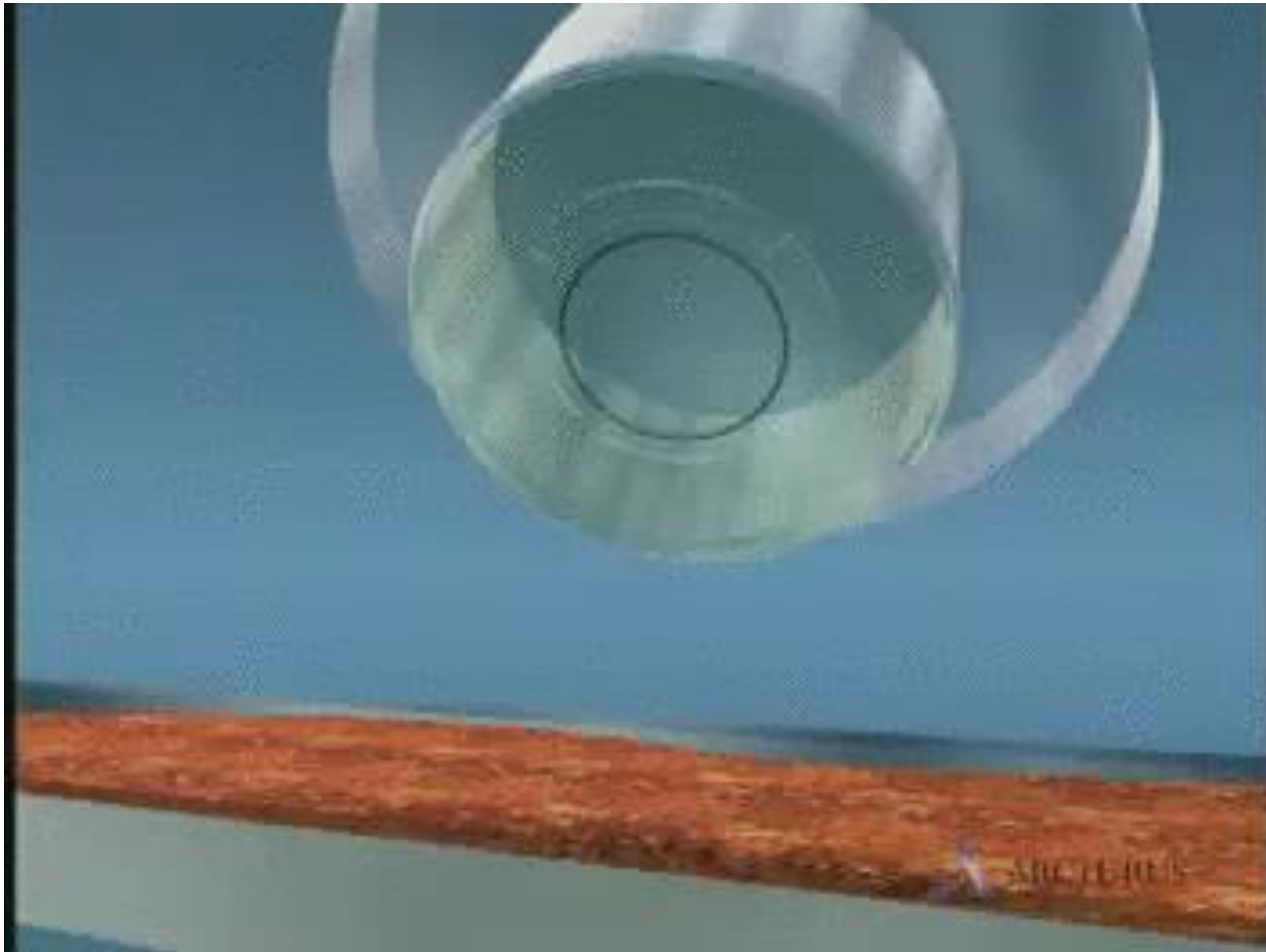
- Inverted bright-field light microscope
- Thermoplastic membrane-mounted cap
- Cap overlies the cells to be microdissected
- Low-power **infrared** (IR) laser beam
- Laser melts a spot of membrane on cells
- Melted membrane attached the cells
- As the plastic cools, the cells are lifted off

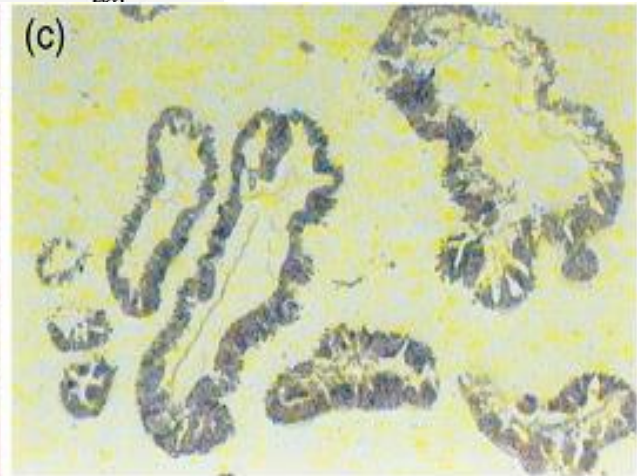
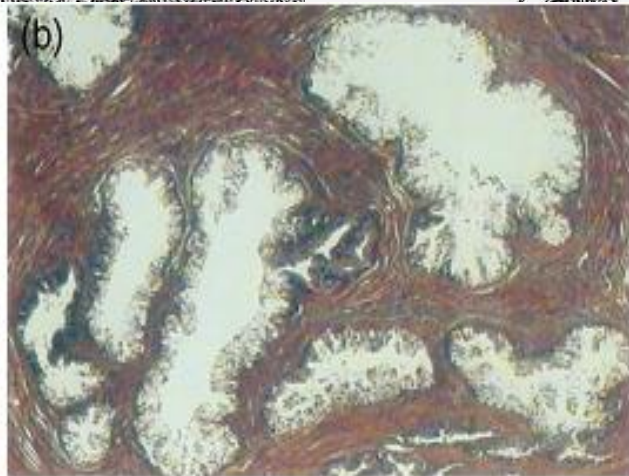
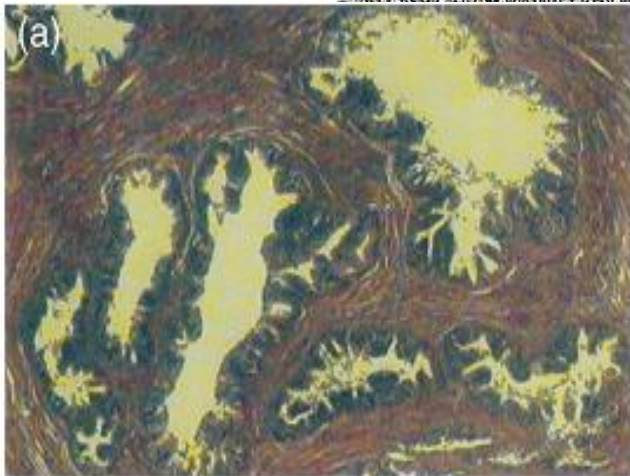
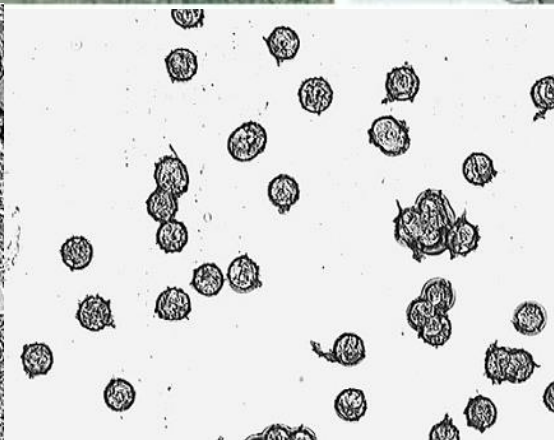
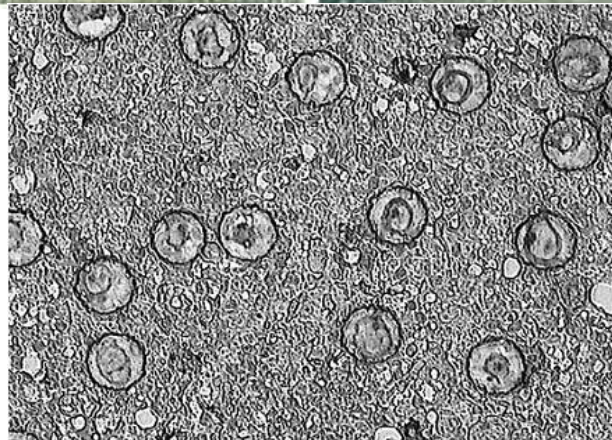
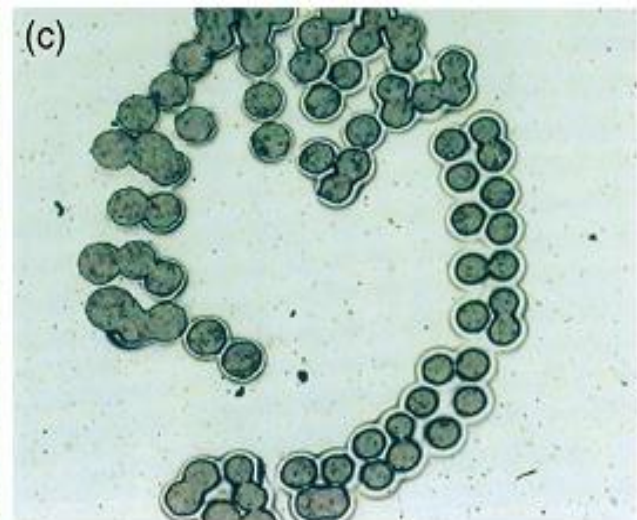
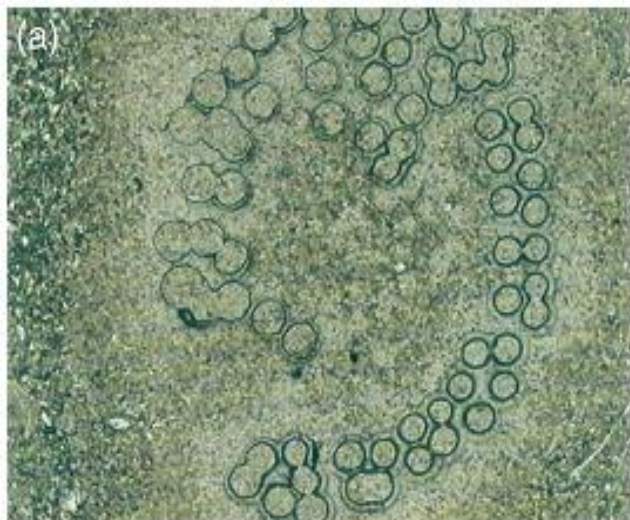
NIH Laser Capture Microdissection





LCM in Action



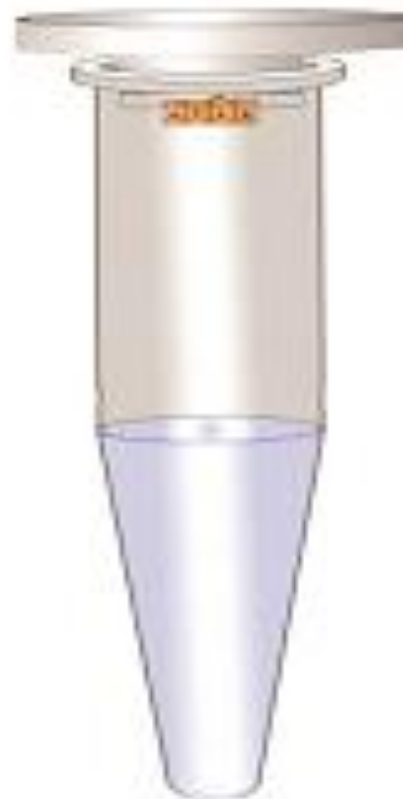




(a)



(b)



(c)



(d)

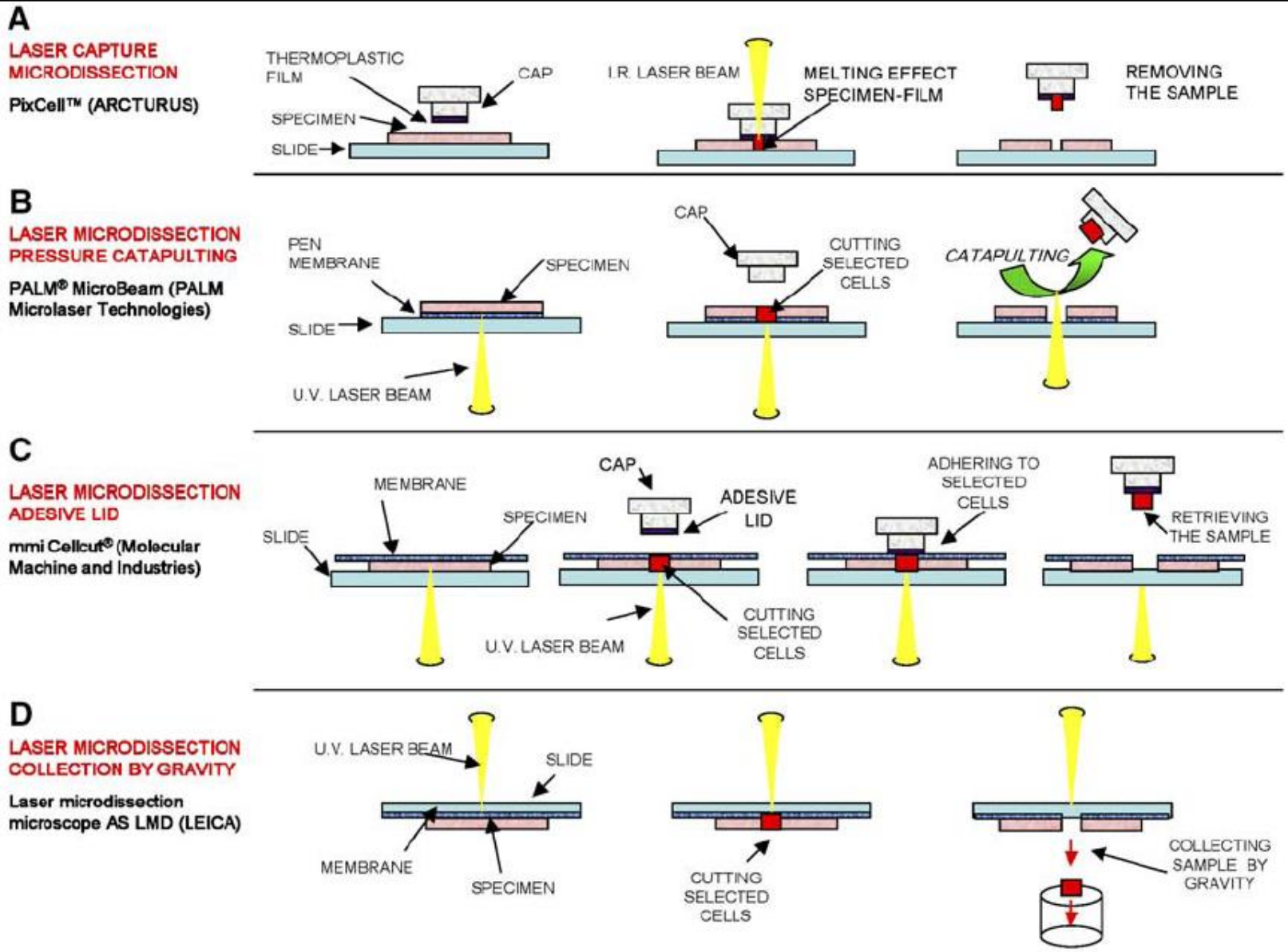


Laser cutting microdissection

- Conceptually different than LCM
- Mostly “**non-contact**”
- Narrow beam of **ultraviolet** (UV) laser
- Laser draw around and cut out the cells
- Dissected cells are collected either by:
 - pressure “catapulting”, OR
 - gravity, OR
 - mechanically acquired by a needle

Pressure “catapulting”







Arcturus PixCell II



A



B

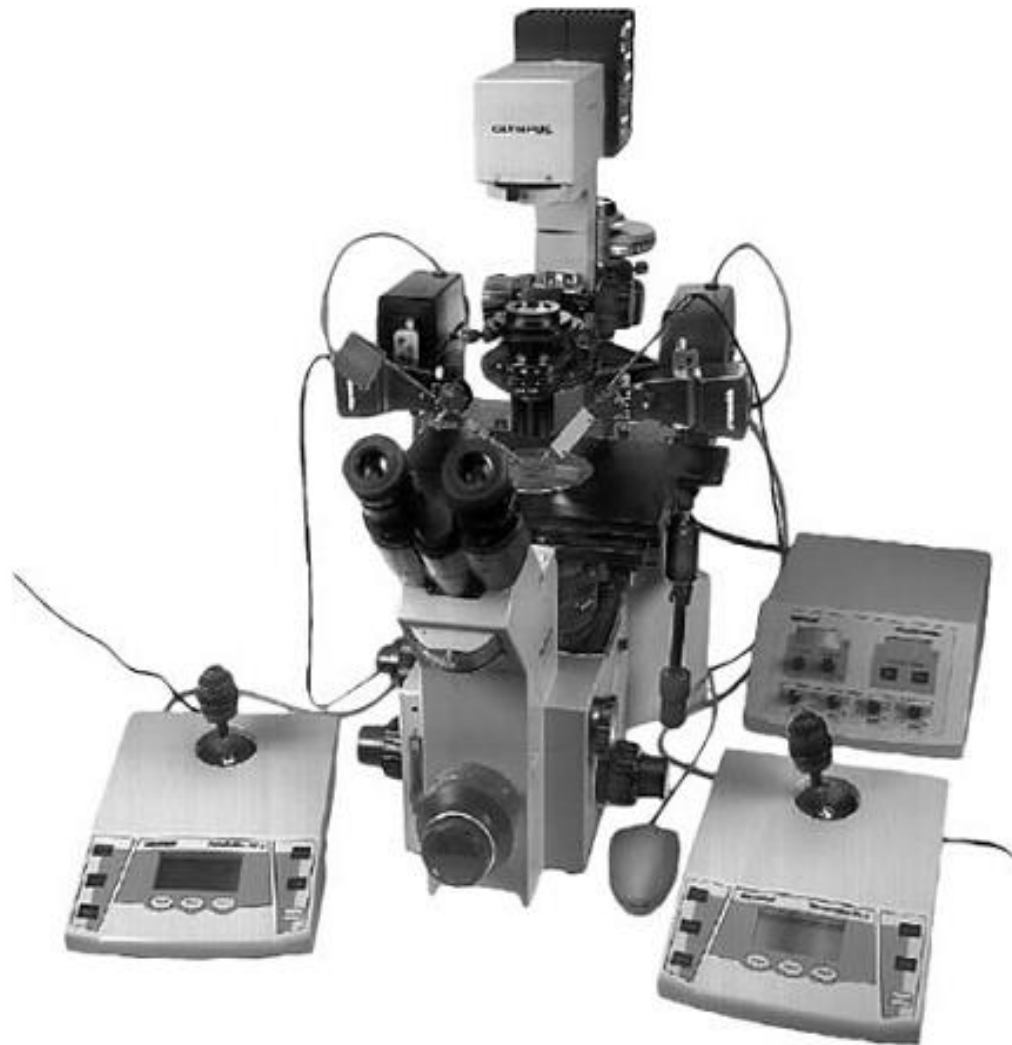


A



B

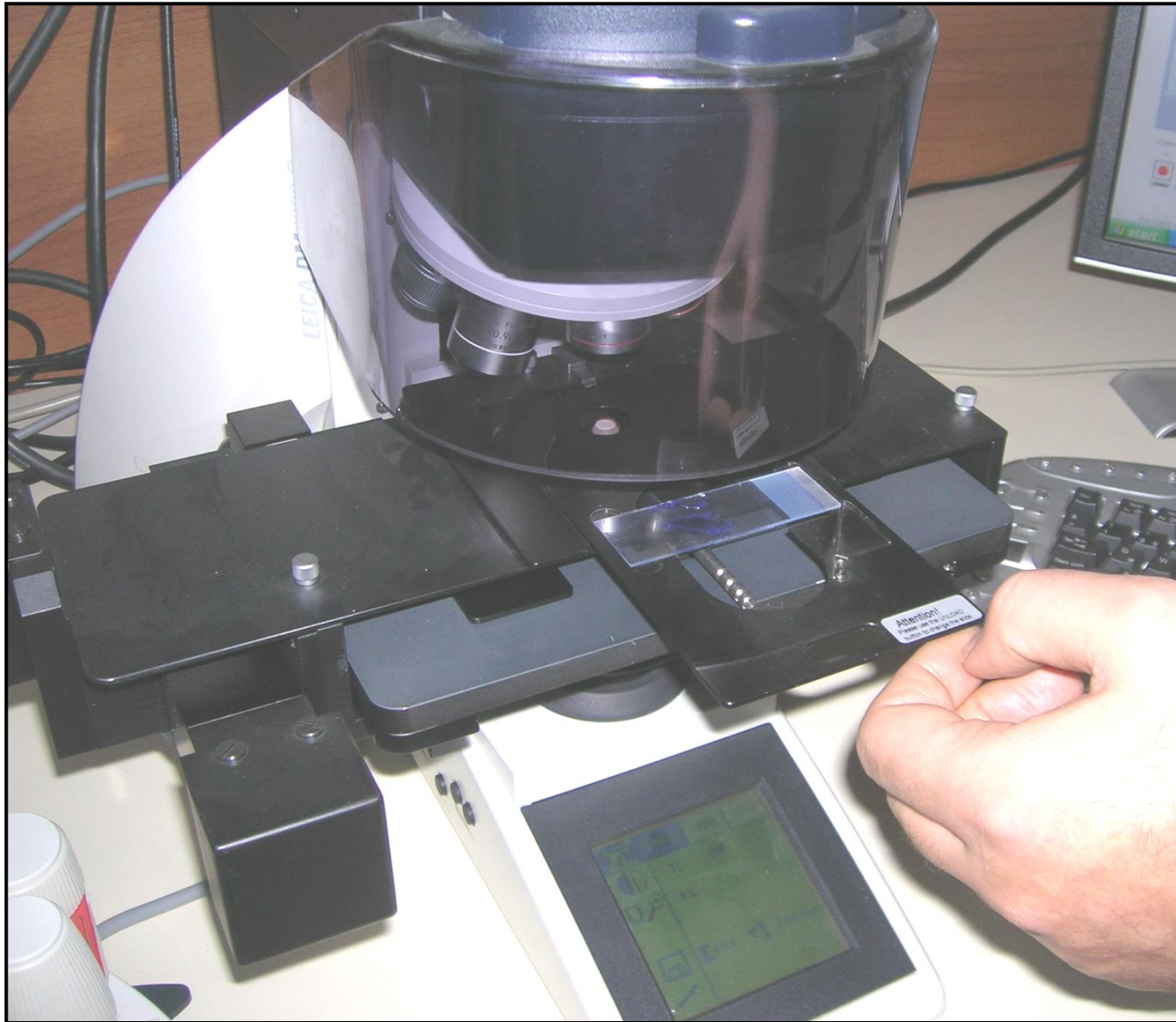




Specimen Preparation

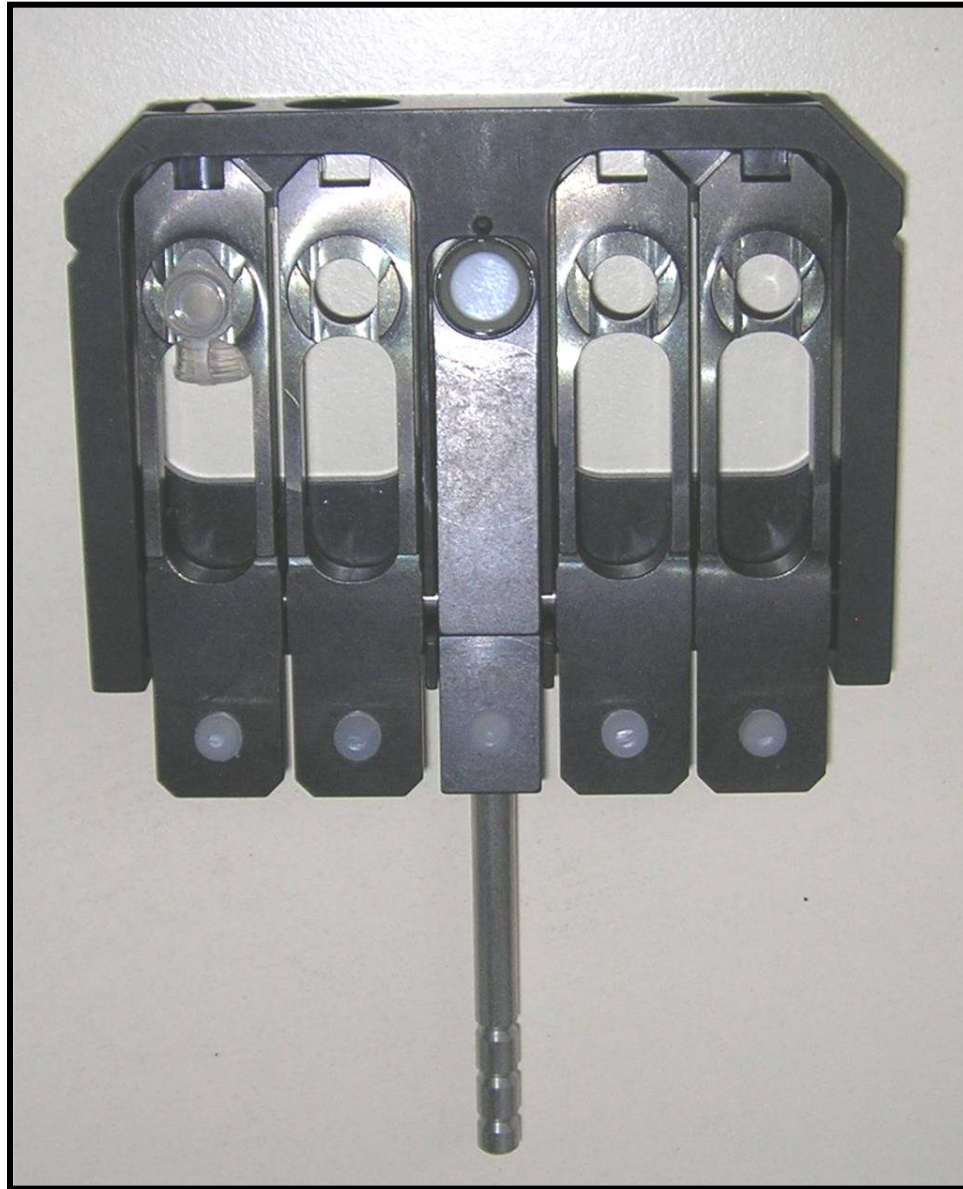
- Any kind of material can be used
- Fresh-frozen tissues or using non-cross linking fixatives are advisable
- Thickness of the section matters (thicker sections require more laser energy)
- Quick staining methods should be used for reducing the possible artifacts
- Adhesive slides may interfere the specimen collection greatly

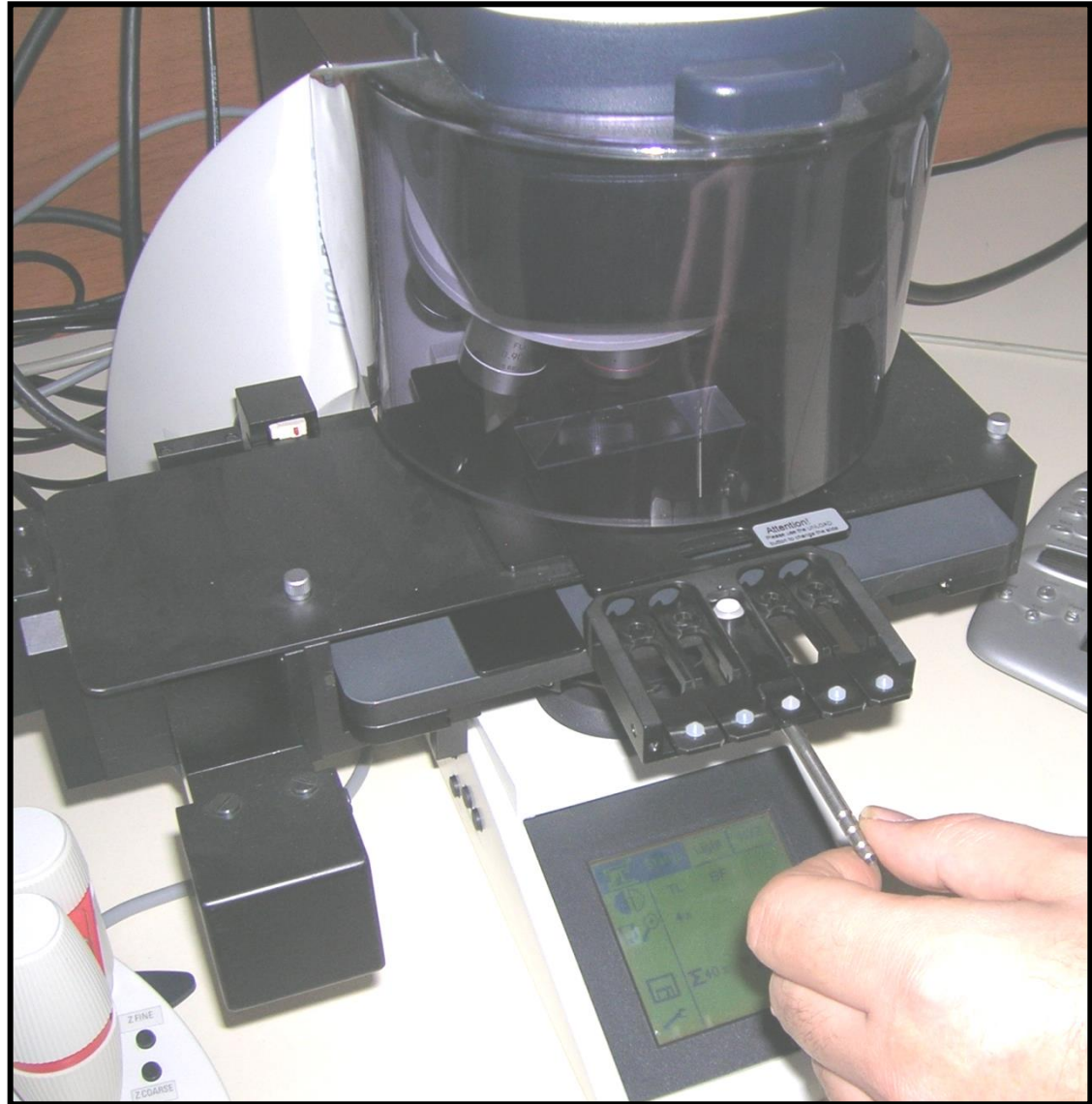
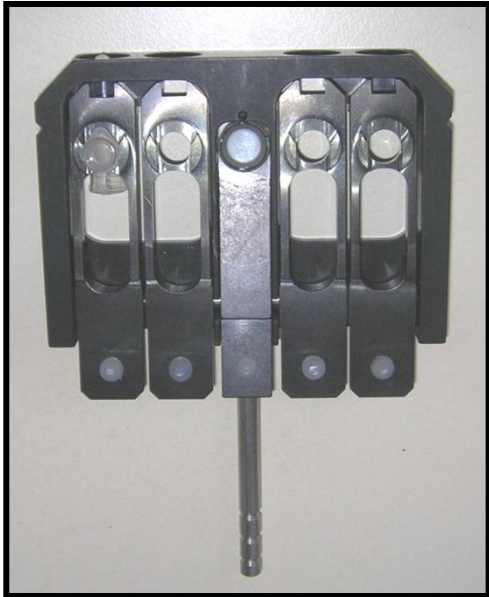












Expected Yields of Nucleic Acids and Proteins

- Each somatic cell contains 7 pg DNA
- 50-100 pg of DNA is enough for PCR
- Total RNA from a cell varies 10-30 pg
- Only 5% of it is mRNA
- Quality of RNA depends on the preservation
- Protein content is tissues specific
- 10-100 times more cells are needed for proteomics

Selected application in Pathology

- Breast cancer
- Colorectal cancer
- Prostate cancer
- Lung cancer

Selected application in Pathology

- Hepatocellular carcinoma
- Neuroblastoma
- Soft-tissue sarcomas
- Lymphoma
- Tuberous sclerosis

DNA-based analyses

- Mutation detection
- Epigenetic alterations
- Loss of Heterozygosity (LOH)
- RFLP
- Comparative genomic hybridization
- Microarrays
- ...

RNA-based analyses

- Gene expression profiles
- Representation difference analysis
- Expressed sequence tag
- Serial analysis of gene expression
- Gene chips
- ...

Protein-based analyses

- Western blotting
- Mass spectrometry
- Peptide sequencing
- Two-dimensional electrophoresis
- Protein microarrays
- ...

microRNA-based analyses

- Posttranscription regulation of genes
- Tissue specific markers
- Developmental lineage signature
- Chemosensitivity and response to treatment
- Diagnostic and prognostic markers
- ...

Future Perspectives

- Developing better tissue preservation methods
- Improved methods on the old FFPE tissues
- Nanomicrodissection (organelles, chromosomes, secretory granules)
- Accelerating various “omics”
- Supporting the individualized patient care

Thank you...